

# A *Drosophila* genetic screen yields allelic series of core microRNA biogenesis factors and reveals post-developmental roles for microRNAs

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## ABSTRACT

Canonical animal microRNAs (miRNAs) are ~22-nt regulatory RNAs generated by stepwise cleavage of primary hairpin transcripts by the Drosha and Dicer RNase III enzymes. We performed a genetic screen using an miRNA-repressed reporter in the *Drosophila* eye and recovered the first reported alleles of fly *drosha*, an allelic series of its dsRBD partner *pasha*, and novel alleles of *dicer-1*. Analysis of *drosha* mutants provided direct confirmation that mirtrons are independent of this nuclease, as inferred earlier from *pasha* knockouts. We further used these mutants to demonstrate in vivo cross-regulation of Drosha and Pasha in the intact animal, confirming remarkable conservation of a homeostatic mechanism that aligns their respective levels. Although the loss of core miRNA pathway components is universally lethal in animals, we unexpectedly recovered hypomorphic alleles that gave adult escapers with overtly normal development. However, the mutant photoreceptor neurons exhibited reduced synaptic transmission, without accompanying defects in neuronal development or maintenance. These findings indicate that synaptic function is especially sensitive to optimal miRNA pathway function. These allelic series of miRNA pathway mutants should find broad usage in studies of miRNA biogenesis and biology in the *Drosophila* system.

**Keywords:** microRNA; Drosha; Pasha; Microprocessor; synapse; Dicer

## INTRODUCTION

microRNAs (miRNAs) are an abundant class of ~22-nt regulatory RNAs derived from endogenous inverted repeat transcripts, which collectively play substantial roles during development and homeostasis (Flynt and Lai 2008). Biogenesis of canonical animal miRNAs follows a conserved strategy involving stepwise cleavage by the Drosha and Dicer RNase III enzymes (Kim et al. 2009). In the *Drosophila* pathway, hairpins within primary miRNA (pri-miRNA) transcripts are recognized in the nucleus by the dsRNA binding domain (dsRBD) protein Pasha, which positions its partner Drosha to cleave ~10 bp into the stem (Han et al. 2006). The Drosha–Pasha complex is often referred to as the “Microprocessor,” whose activity yields ~55–70-nt pre-

miRNA hairpins (Denli et al. 2004; Gregory et al. 2004; Han et al. 2004). These are exported to the cytoplasm by Exportin-5/Ran-GTP (Yi et al. 2003; Bohnsack et al. 2004; Lund et al. 2004) and cleaved again by Dicer-1 in complex with its dsRBD partner Loqs (Forstemann et al. 2005; Saito et al. 2005). One strand of each resulting duplex is preferentially loaded into the miRNA effector Argonaute1 (AGO1) (Forstemann et al. 2007; Tomari et al. 2007; Okamura et al. 2009), which partners with GW182 (Gawky) to repress target transcripts (Liu et al. 2005; Rehwinkel et al. 2005; Eulalio et al. 2008). As in other animals, functional *Drosophila* miRNA targets generally exhibit limited complementarity and frequently pair only to positions 2–8 of the miRNA (Lai 2002; Ruby et al. 2007b), also known as the “seed” region (Bartel 2009). Productive regulatory interactions usually lead to target deadenylation and degradation, although translational inhibition and/or target relocalization also contribute to miRNA-mediated silencing (Lai and Posakony 1997; Lai et al. 1998; Behm-Ansmant et al. 2006; Zdanowicz et al. 2009).

In addition to the canonical Drosha–Dicer pathway, several alternate pathways for miRNA biogenesis have been

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documented (Yang and Lai 2011). For example, mirtrons are short hairpin introns that are spliced and debranched to generate pre-miRNA hairpins, thereby bypassing the need for Drosha cleavage (Okamura et al. 2007; Ruby et al. 2007a). Variant mirtrons with 5' or 3' tails rely on splicing to generate one hairpin end but require other exonucleases to trim their tails to permit nuclear export and dicing (Babiarz et al. 2008; Flynt et al. 2010). Recently, vertebrate *mir-451* was recognized to be Drosha-dependent but Dicer-independent, and instead matures via the catalytic activity of Ago2 (Cheloufi et al. 2010; Cifuentes et al. 2010; Yang et al. 2010). Nevertheless, the majority of miRNAs proceed through the canonical biogenesis pathway.

Most studies of miRNA biogenesis have relied on biochemical approaches and usage of RNA interference to reduce the activity of candidate factors, even in traditionally genetic systems such as *Drosophila* and *Caenorhabditis elegans* (Eulalio et al. 2007; Parry et al. 2007; Zhou et al. 2008). Consequently, there are few mutant alleles available for many key miRNA pathway genes. For example, in flies, there are only two alleles of *dicer-1* (Lee et al. 2004; Berdnik et al. 2008), two alleles of *pasha* (Berdnik et al. 2008; Martin et al. 2009), and no mutant alleles described for *drosha*. The paucity of mutants limits the flexibility of genetic analysis for miRNA pathway function, especially in ways that would otherwise be facilitated by the availability of allelic series. For example, an extensive collection of *dicer-2* alleles includes mutants that permit genetic separation of the role of this enzyme for siRNA genesis from its role in siRNA loading into AGO2 complex (Lee et al. 2004; Tomari et al. 2004; Lim et al. 2008).

Forward genetic screening offers a strategy to obtain mutants in an unbiased fashion and has the potential to identify new pathway components and accessory factors. In the present study, we describe a *Drosophila* genetic screen designed to identify mutants that affect the biogenesis or activity of canonical miRNAs. The efficacy of this screen was demonstrated by recovery of the first reported alleles of *Drosophila drosha*, the first point mutations of *pasha*, and several novel alleles of *dicer-1*. We use these alleles to demonstrate mutual cross-regulation of Drosha and Pasha levels, previously characterized only in cultured cells (Han et al. 2009; Kadener et al. 2009; Triboulet et al. 2009), for the first time in the intact animal. Finally, the availability of allelic series of these genes allowed us to access post-developmental roles of the miRNA pathway. In particular, we find that mild reduction of global miRNA levels leads to a synaptic transmission defect without overtly affecting development or neuronal maintenance.

## RESULTS

### Development of a simple in vivo visual reporter for miRNA pathway activity

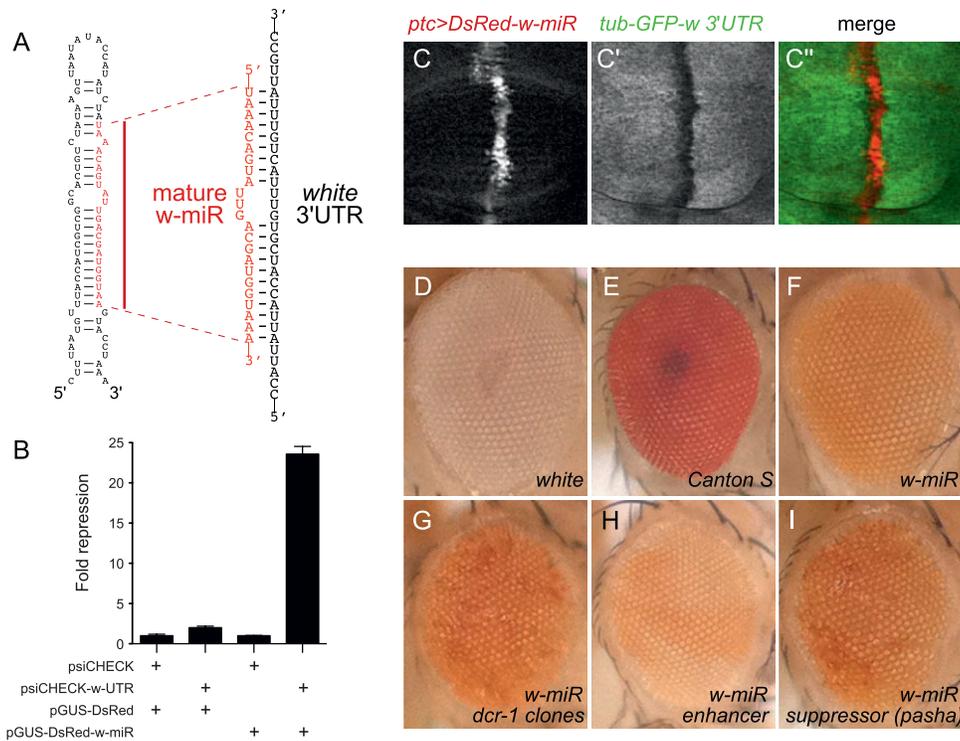
To facilitate large-scale genetic identification of loci required for miRNA biogenesis or function, we designed a system to

assay miRNA activity via simple visual inspection of eye pigmentation. A similar strategy was previously used successfully to identify genes involved in RNAi in *Drosophila* (Lee et al. 2004; Kim et al. 2007; Lim et al. 2008; Lee et al. 2009; Marques et al. 2010). To begin, we reprogrammed the *mir-6-1* backbone (Chen et al. 2007) to generate an miRNA that targets a single location in the endogenous 3' UTR of the *white* gene (*w-miR*) (Fig. 1A). To encourage miRNA-like biogenesis and target regulation (Forstemann et al. 2007; Czech et al. 2009; Okamura et al. 2009; Ghildiyal et al. 2010), we designed *w-mir* to have a central bulge in its pre-miRNA hairpin and mismatches at positions 10–12 relative to the target site (Fig. 1A). This miRNA was placed downstream from *DsRed* under control of GUS regulatory sequences (Brodsky et al. 2000); this hybrid element combines the GMR enhancer for specific expression in the developing eye and UAS sites for optional Gal4-mediated control. *GUS-DsRed-w-miR* was carried in a *P*-element transformation vector containing a standard *mini-white* construct bearing the endogenous *white* 3' UTR.

To assay the regulatory activity of *w-miR*, we first performed sensor assays in S2 cells. Cotransfection of *GUS-DsRed-w-mir* and *ub-Gal4* specifically repressed a *Renilla* luciferase reporter bearing the *white* 3' UTR, compared with controls (Fig. 1B). We next performed more physiological tests using third instar wing imaginal discs. We prepared a *tub-GFP-white* 3' UTR transgene, which is expressed ubiquitously in the animal. When introduced into a background carrying *GUS-DsRed-w-mir* and *ptc-Gal4*, thus activating DsRed and *w-miR* along the anterior–posterior compartment boundary, we correspondingly observed cell-autonomous repression of the *GFP-white* 3' UTR sensor (Fig. 1C). Therefore, *w-miR* was able to silence transcripts bearing a single bulged site in the *white* 3' UTR in vivo.

Finally, we assayed the capacity of *w-miR* to suppress pigmentation. When placed in a background that is mutant for the endogenous *white* locus, *GUS-DsRed-w-mir* (hereafter abbreviated as *w-mir*) is autotargeting in the eye, since it generates both the *white* gene product from the *mini-white* transformation marker in the vector backbone and an miRNA that silences this transcript. In principle, if such suppression were sufficiently robust, we may not have been able to score insertions based on standard rescue of eye pigment; we therefore included a DsRed selection marker. It turns out that multiple independent *w-mir* insertions were easily isolated, and these bore a range of eye colors as is typical for random *P* integration. On the whole, these independent transgenes had mild levels of pigment (Fig. 1F, cf. wild-type in Fig. 1E and the *white*-null mutant in Fig. 1D).

To test if *w-miR* was indeed responsible for this apparent decrease in eye pigment, we generated eye clones for a null mutation of *dcr-1* (Lee et al. 2004) in the background of *w-miR*. The resulting eyes appeared mosaic and had patches of tissue that were small and rough, as is characteristic of *dcr-1* clonal tissue. Importantly, these mutant regions



**FIGURE 1.** A synthetic miRNA directed against *white* is functional in vivo and a useful screening tool for miRNA biogenesis factors. (A) Predicted secondary structure of w-miR and its predicted interaction with the target sequence in the *white* 3' UTR. The predicted mature strand is indicated in red. (B) S2 cells were transfected with the indicated plasmids, together with ub-gal4. Values represent fold repression of luciferase activity relative to empty psiCHECK vector. (C) Wing disc sensor assay. w-miR expression driven by *ptc*-gal4 (C) results in repression of tub-GFP-w-3'UTR sensor (C') and merge, C''). (D) A *white* eye. (E) A wild-type (*Canton S*) eye. (F) Eye of an animal with pGUS-DsRed-w-miR transgene. (G) Clones of *dcr-1*[Q1147X] in the background of GMR-w-miR result in a mosaic eye with darker patches. (H) Example of presumed enhancer of miR-mediated silencing from our screen (gene unknown). (I) Example of suppressor of miR-mediated silencing from our screen (*pasha*[23D2], which is a newly isolated null mutant of *pasha* that results in the premature truncation of Pasha at R308).

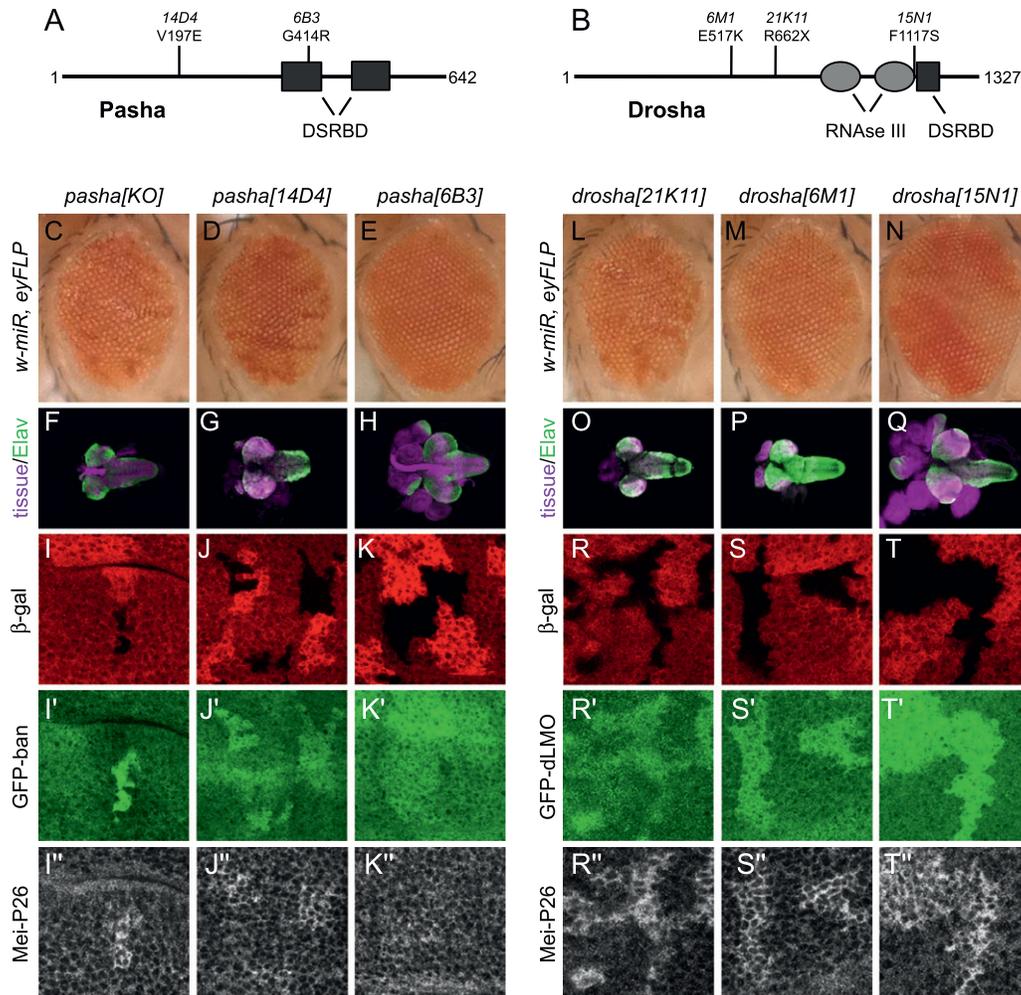
exhibited darker red pigmentation than surrounding tissue, indicating that *dcr-1*, and by inference the miRNA pathway, was required for w-miR-mediated suppression of eye pigmentation (Fig. 1G). We concluded that w-miR is a suitable reagent with which to carry out forward genetic screening for loci required for the miRNA pathway.

### Genetic screening using w-miR recovers novel Microprocessor mutations

We used the suppression of eye pigment by w-miR as the basis of F<sub>1</sub> genetic screens of the right arms of chromosome 2 and 3; screening of other chromosome arms is ongoing. We scored for eyes exhibiting mosaicism of eye color, then backcrossed and retested these for germline transmission of mutations and reproducibility of pigment modification. In principle, clonal changes in eye color can result in lighter patches (in which case, the mutation potentially enhances miRNA activity) or darker pigment (in which case, the mutation potentially suppresses miRNA activity). Examples of enhancers and suppressors of w-miR are shown in Figure 1H and 1I, respectively. Of course, it is also possible that clonal changes in eye color were due to mutations in the pigment

biosynthetic pathway. Therefore, we performed a secondary test for the dependence of pigment modification on w-miR. Instead of using the *eyflp* transgene to generate clones, we used a chromosome with *ey-gal4* and *UAS-FLP*, both of which are carried on transgenes that use *white*<sup>+</sup> as a selectable marker (Stowers and Schwarz 1999). Any mutant alleles that gave eye color mosaicism in this background were discarded because their effect on pigmentation is not w-miR-dependent. A full description of mutants isolated from these screens will be reported elsewhere (P Smibert and EC Lai, in prep.).

Two suppressor mutations recovered from chromosome 3R, *Su6B3* and *Su14D4*, failed to complement each other and also failed to complement a previously described *pasha* deletion allele, *pasha*[KO] (Martin et al. 2009). We isolated genomic DNA from larvae trans-heterozygous for these mutations against *pasha*[KO], and amplified and sequenced the mutant *pasha* locus. Both alleles had missense mutations in the coding exons of *pasha* (Fig. 2A,D,E). Three suppressor mutations on chromosome 2R—*Su6M1*, *Su15N1*, and *Su21K11*—also exhibited eye mosaicism with patches of more pigment (Fig. 2L–N). All three failed to complement *Df(2R)exel6055*, which uncovers 33 genes (<http://flybase.org/>), including *droscha*. Sequencing of DNA from



**FIGURE 2.** Identification and characterization of novel Microprocessor alleles. (A) Molecular lesions of *pasha* alleles. *pasha*[*KO*] is a complete locus deletion that has been described previously (Martin et al. 2009). (B) Molecular lesions of *drosha* alleles. (C–K) Phenotypic characterization of *pasha* alleles: (C–E) Adult eyes with clones of the indicated genotype in the background of GMR-*w-miR*. Note that the *pasha*[*KO*] allele itself has a *w*[+] transgene and is included only for comparison of morphology, not pigmentation. (F–H) Third instar disc clusters of indicated alleles *in trans* with *pasha*[*KO*] stained with Elav (green) and a tissue marker (Phalloidin, [*KO*] and [6B3]; or Hoechst, 33352-[14D4], purple). (I–K) Mitotic clones of indicated alleles marked by the absence of  $\beta$ -gal (red) and stained for GFP-ban sensor (green, I'–K') and Mei-P26 (grayscale, I''–K''). (L–T) Phenotypic characterization of *drosha* alleles: (L, M) Adult eyes with clones of the indicated genotype in the background of GMR-*w-miR*. (O–Q) Third instar disc clusters of indicated alleles *in trans* with *Df(2R)exel6055* stained with Elav (green) and a Hoechst 33352 (purple). (R–T) Mitotic clones of indicated alleles marked by the absence of  $\beta$ -gal (red) and stained for GFP-ban sensor (green, R'–T') and Mei-P26 (grayscale, R''–T'').

larvae *trans*-heterozygous for *Df(2R)exel6055* and each mutation revealed that all had nonsynonymous mutations in *drosha*-coding exons (Fig. 2B). One of these alleles, *Su21K11*, carried an R662X mutation that results in premature termination of the protein before the RNase III domains and is therefore expected to be a null mutant. This mutant behaves genetically as a null, as homozygotes are indistinguishable from animals *trans*-heterozygous for this allele and *Df(2R)exel6055* (data not shown). To our knowledge, these represent the first reported mutant alleles of *drosha* in *Drosophila*.

These Microprocessor mutants were all recovered on the basis of clonal derepression of *white*<sup>+</sup> function in the eye, but different alleles had distinct effects on eye morphology. Eyes

bearing null clones for either *pasha* or *drosha* were smaller and rough, with disorganized or absent ommatidia (Fig. 2C, L). Eyes bearing clones of *pasha*[14D4] or *drosha*[6M1] were slightly less phenotypically severe than their respective null mutants (Fig. 2D, M). In contrast, clones for *pasha*[6B3] or *drosha*[15N1] appeared morphologically normal, with no obvious defects in ommatidial size or organization (Fig. 2E, N). Additionally, clones of these mutants occupied ~50% of the eye, compared with the stronger mutants whose clones occupied much less. This indicated a growth disadvantage for strong *pasha* and *drosha* alleles that was absent from the corresponding weak alleles.

To assess the overall developmental consequence of flies mutant for our novel alleles, we examined third instar brain

and imaginal disc clusters. To rule out potential contributions from second site mutations, as well as to provide direct comparison of allelic strengths, we placed each *pasha* or *drosha* allele *in trans* to the *pasha*[*KO*] or *Df*(2*R*)*exel6055* deletions, respectively. For simplicity, we refer to these *trans*-heterozygous allelic combinations by the allele of interest. Homozygous *pasha*[*KO*] animals were used because these animals are rescued to viability with a genomic rescue transgene (Martin et al. 2009).

As described previously (Martin et al. 2009), *pasha*[*KO*] animals form small third instar larvae that are almost completely devoid of imaginal discs (Fig. 2F). These animals die as larvae or poorly formed early pupae. Some larvae develop rudimentary eye discs, but most animals lack any obvious imaginal discs. *pasha*[*14D4*] mutants have rudimentary eye discs that stain for the neural marker Elav (Fig. 2G), and these animals die as early pupae. In contrast, *pasha*[*6B3*] animals have a virtually wild-type complement of imaginal discs surrounding a normally sized brain and go on to die as pharate adults (Fig. 2H). Similar to *pasha*-null animals, *drosha*[*21K11*] animals die as third instar larvae or poorly formed early pupae. At the third larval instar, these animals are almost completely lacking imaginal disc structures (Fig. 2O). *drosha*[*6M1*] animals exhibit an intermediate phenotype with respect to imaginal tissues at the third larval instar, with rudimentary Elav positive eye discs, but no other imaginal tissues (Fig. 2P). In contrast, *drosha*[*15N1*] animals have a full complement of imaginal discs and look morphologically indistinguishable from wild type (Fig. 2Q). While *trans*-heterozygotes of *drosha*[*15N1*] over the deletion die as pharate adults, *drosha*[*15N1*] homozygotes occasionally survive to adulthood.

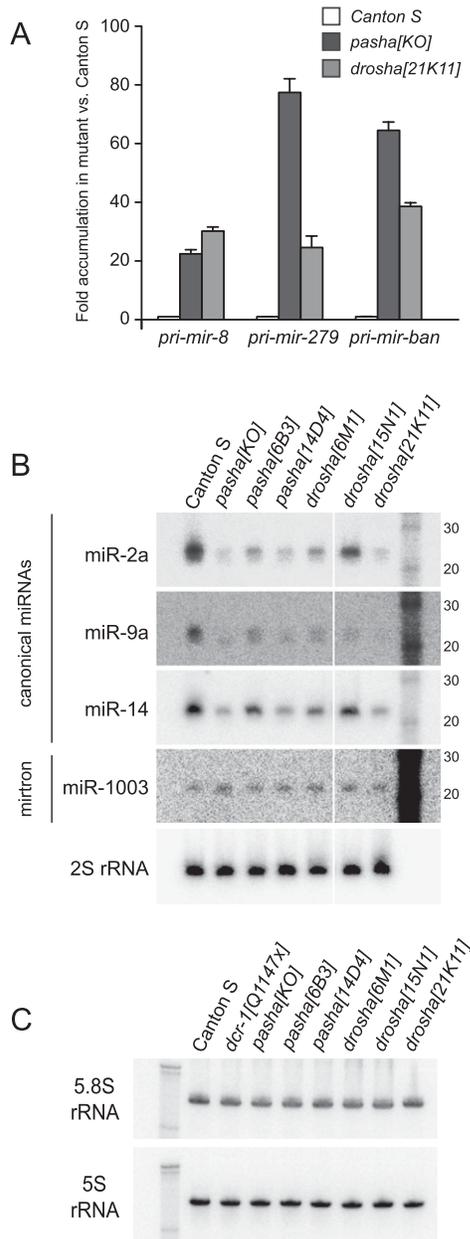
### Differential defects in miRNA biogenesis and function in Microprocessor mutants

It was unexpected that screening for failure to repress a miRNA-sensitive reporter could yield *pasha* and *drosha* alleles that could survive to adulthood, as the miRNA pathway is universally essential for viability in all animals. We therefore conducted a detailed series of functional assays of miRNA biogenesis and function on our allelic series of *pasha* and *drosha* mutants. We first generated mitotic clones in the wing imaginal disc and assayed for the levels of previously described sensors of miRNA activity. We used *tub*-*GFP*-*bantam* and *tub*-*GFP*-*dLMO*, which are ubiquitously expressed *GFP* transgenes driven by the  $\beta$ -tubulin promoter followed by two copies of a sequence complementary to the *bantam* miRNA, or the 3' UTR of the miRNA-regulated gene *dLMO* (Brennecke et al. 2003; Bejarano et al. 2010). Additionally, we stained for endogenous Mei-P26 protein, a miRNA target that is derepressed in *dcr-1* and *ago1* mutant clones (Herranz et al. 2010). In all alleles examined, homozygosity for the Microprocessor mutations resulted in derepression of both the *GFP* sensors

and Mei-P26 (Fig. 2I–K,R–T). In the case of *pasha* mutations, the level of derepression of both *GFP*-*bantam* and Mei-P26 correlated with allele strength with strong derepression for the null mutant and only very mild derepression in the weak *pasha*[*6B3*] clones (Fig. 2I–K). However, despite the difference in phenotypic severity of the *drosha* alleles examined at the level of morphology, no obvious differences in the level of derepression of either *GFP*-*dLMO* or Mei-P26 were observed (Fig. 2R–T). In other words, these weak *drosha* alleles nevertheless exhibited strong derepression of artificial and even endogenous miRNA targets.

The roles of *Drosha* in miRNA biogenesis, rRNA maturation, and mRNA cleavage have been extensively studied *in vitro* using biochemistry and knockdowns in cultured cells, but *in vivo* studies are comparably limited. We took the opportunity to test the requirements for *Drosha* in RNA metabolism using these first-available *Drosophila drosha* mutants. The best characterized role for *Drosha* is to cleave pre-miRNAs from primary miRNA transcripts (Lee et al. 2003; Denli et al. 2004; Gregory et al. 2004; Han et al. 2004; Landthaler et al. 2004). A subset of miRNAs is produced from *Drosha*-independent substrates, including mirtrons, which are short hairpin introns that use splicing to generate the pre-miRNA (Okamura et al. 2007; Ruby et al. 2007a). Our studies of a *pasha* knockout verified that this *Drosha* cofactor was required for *in vivo* maturation and function of canonical miRNAs but not mirtrons (Martin et al. 2009). Here, we demonstrate the direct involvement of the *Drosha* nuclease for canonical miRNA biogenesis in the animal. qRT-PCR assays of 5–6-d-old third instar larvae revealed strong accumulation of primary miRNA transcripts in *drosha*[*21K11*] animals, comparable to *pasha*[*KO*] animals isolated in parallel (Fig. 3A). Northern analysis of the whole panel of Microprocessor mutants showed that levels of mature miR-14, miR-2a, and miR-9a were decreased in all cases, whereas mirtron-derived miR-1003 was unaffected in any of the mutants (Fig. 3B).

Notably, for both *pasha* and *drosha* mutants, the level of reduction in mature miRNAs correlated with the strength of developmental and viability defects observed. The strongest loss of mature miRNA species was observed for the null mutations, *pasha*[*KO*] and *drosha*[*21K11*], followed by the intermediate strength mutations, *pasha*[*14D4*] and *drosha*[*6M1*]. The weak hypomorphic mutations, *pasha*[*6B3*] and *drosha*[*15N1*], had only modest reductions in mature miRNA levels, consistent with the modest phenotypes associated with their homozygosity (Figs. 2C–H,I,L–Q, 3B). Given this correlation, it was surprising that we observed strong derepression of multiple miRNA sensors and even of the endogenous miRNA target Mei-P26 in mutant clones of *drosha*[*15N1*], a weak allele that exhibits only subtle effects on absolute mature miRNA levels. We hypothesize that certain readouts and/or targets of miRNA-mediated repression are more sensitive than others, a ge-



**FIGURE 3.** Effects of Microprocessor mutants on small RNA maturation. (A) Accumulation of pri-miRNA transcripts in null Microprocessor mutants. Total RNA was extracted from Canton S, homozygous *pasha*[KO], or trans-heterozygous *drosha*[21K11]/*Df(2R)exel6055* animals and assayed by qRT-PCR. The y-axis represents the fold up-regulation of the indicated miRNA relative to the Canton S sample and standard deviation. (B) Mature miRNA levels in Microprocessor mutants. Total RNA was extracted from third instar larvae of the indicated genotypes. For the *pasha* alleles, indicated alleles were *in trans* with *pasha*[KO]. For the *drosha* alleles, indicated alleles were *in trans* with *Df(2R)exel6055*. The small RNA blot was sequentially probed as indicated. 2S rRNA was used as a loading and transferring control. (C) Ribosomal RNA maturation is unaffected in Microprocessor mutants. RNA from the same samples was run simultaneously on two gels, transferred, and probed for 5S and 5.8S rRNA.

netic situation that can be appreciated by availability of an allelic series. We revisit the biological implications of this below in this study.

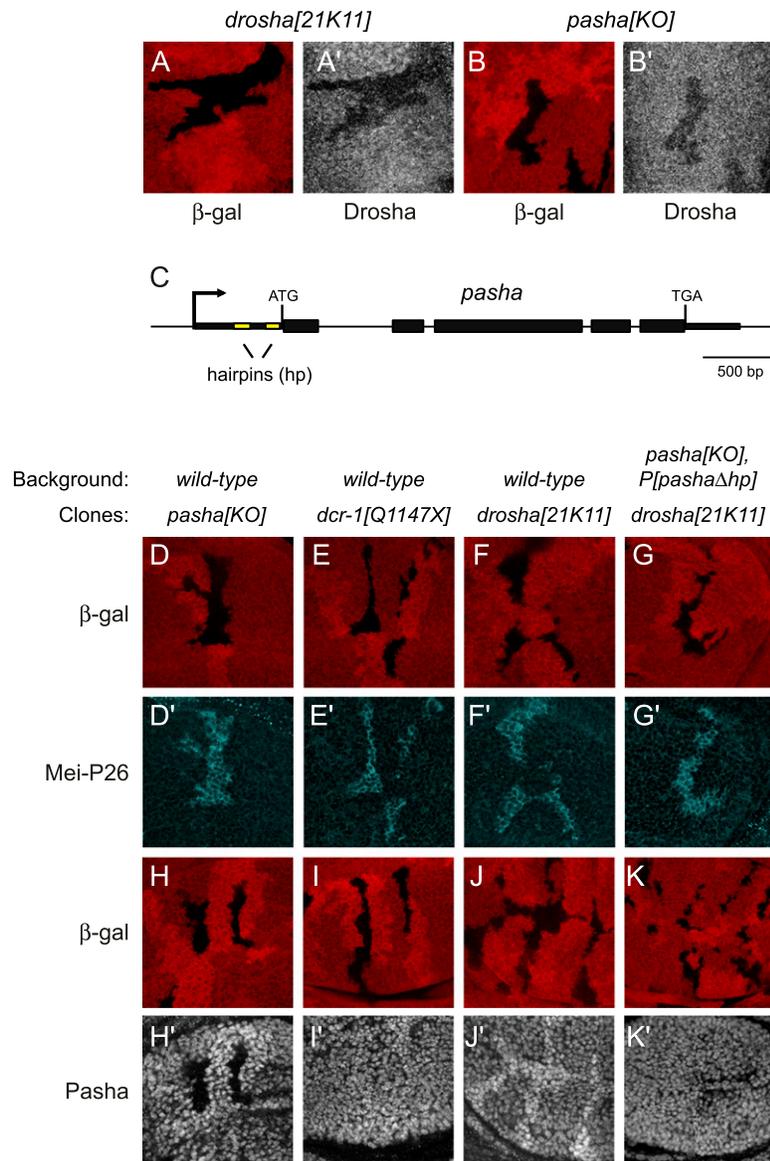
Drosha was initially identified as an RNase III enzyme required for maturation of ribosomal RNAs. Antisense inhibition of Drosha resulted in accumulation of 12S and 32S rRNA species, interpreted as a partial block in rRNA processing (Wu et al. 2000). Subsequently, loss of the Microprocessor accessory factors, the DEAD-box helicases p68 and p72, was reported to decrease 5.8S rRNA maturation (Fukuda et al. 2007). We analyzed 5.8S rRNA levels across our panel of Microprocessor mutants at the third larval instar, which are presumably depleted of stores of maternal protein inherited 5–6 d earlier. However, we did not observe any obvious differences from control Canton S animals (Fig. 3C). The absence of rRNA biogenesis defects in *drosha* null flies parallels an analysis from *Drosophila* knockout mouse thymocytes, which indicated no defects in rRNA levels (Chong et al. 2008).

### In vivo demonstration of cross-regulation of the Microprocessor

In addition to its major role in nuclear miRNA biogenesis, Drosha also cleaves certain mRNA hairpins. Its most well-characterized mRNA target encodes DGCR8, the mammalian ortholog of Pasha (Han et al. 2009; Triboulet et al. 2009). Reciprocally, DGCR8/Pasha proteins are required for Drosha stability, thus fulfilling a post-transcriptional and post-translational homeostatic loop that tunes appropriate levels of these core Microprocessor components (Han et al. 2009). Studies in S2 cell culture showed that the capacity of Drosha to cleave and down-regulate *pasha* transcript is conserved in *Drosophila* (Han et al. 2009; Kadener et al. 2009).

We used our Microprocessor mutant alleles to perform strict genetic tests of the Microprocessor cross-regulatory loop in the animal. Mitotic clones of *drosha*[21K11] showed derepression of both an endogenous miRNA target and a sensor for miRNA activity (Fig. 2R) and a strong reduction of Drosha (Fig. 4A). Unfortunately, the available Drosha antibody (Denli et al. 2004) is not entirely specific in tissue staining but exhibits strongly decreased immunoreactivity in *drosha*-null clones. Mitotic clones of *pasha*[KO] also showed derepression of both an endogenous miRNA target and a sensor for miRNA activity (Fig. 2I). Furthermore, *pasha*[KO] clones exhibited strongly reduced Drosha staining equivalent to *drosha* mutant clones (Fig. 4B), indicating that *Drosophila* Pasha is required for Drosha stability in the animal.

Knockdown of *drosha* in S2 cells increases *pasha* transcript levels (Han et al. 2009), but the effect on Pasha protein was unknown. We generated mitotic clones of null mutants of *pasha* (Martin et al. 2009), *dicer-1* (Lee et al. 2004), and *drosha* (this work), all of which derepress miRNA sensors and endogenous miRNA targets (Figs. 2, 4D–F), and assayed



**FIGURE 4.** In vivo cross-regulation of Microprocessor components in *Drosophila*. In all cases, mutant clones are marked by the absence of β-gal (stained in red). (A) A clone for *drosha[21K11]* has strongly reduced Drosha staining (A'). (B) A *pasha[KO]* clone also has strongly reduced Drosha staining (B'). (C) Schematic representation of *pasha* locus demonstrating position of 5'-UTR hairpins. (D–G) Clones of *pasha[KO]* (D), *dcr-1[Q1147X]* (E), and *drosha[21K11]*, either in a wild-type background (F) or in a background homozygous for *pasha[KO]* rescued by two copies of a *pasha* genomic rescue fragment lacking the two hairpins in the 5' UTR (G) exhibit derepression of Mei-P26 (D'–G'), indicating loss of miRNA pathway activity. (H–K) Clones of the same genotypes as D–G have different effects on Pasha levels (H'–K'). (H) Clones of *pasha[KO]* have no detectable Pasha staining. (I) Pasha levels are unaffected in *dcr-1[Q1147X]* clones. (J) Pasha protein is elevated in clones of *drosha[21K11]*. (K) In a genetic background homozygous for the *pasha[KO]* allele but rescued with two copies of a *pasha* genomic rescue transgene lacking the two hairpins in the 5' UTR, clones for *drosha[21K11]* do not have altered Pasha levels.

Pasha protein levels. As expected, clones homozygous for a null *pasha* mutant lack Pasha completely (Fig. 4H). Clonal loss of Dcr-1 had no discernible effect on Pasha levels (Fig. 4I), demonstrating that Pasha levels are not significantly affected by global reduction of miRNAs. However, loss of Drosha,

which also globally reduces miRNAs and miRNA pathway activity (Figs. 2R, 4F), resulted in a cell-autonomous increase in Pasha (Fig. 4J). Since Pasha protein was not up-regulated in *dcr-1* clones, its up-regulation in *drosha* clones is miRNA-independent and instead consistent with direct cleavage of *pasha* transcript by Drosha.

We performed a stringent test of this notion by introducing a mutated *pasha* genomic construct into the *pasha* deletion background, such that the only source of *pasha* was transgenically expressed under endogenous transcriptional control. We identified two *pasha* 5'-UTR hairpins that generated short RNA reads consistent with a low level of miRNA-like biogenesis (Supplemental Fig. 2) and deleted these from the construct. This transgene was inserted onto chromosome 3L and recombined with *pasha[KO]*. We then introduced this recombinant third chromosome into the FRT42D *drosha[21K11]* background and generated negatively marked *hs-FLP* clones. In this experiment, despite the loss of Drosha and the derepression of an endogenous miRNA target (Fig. 4G), the level of Pasha protein was unchanged (Fig. 4K). This confirms that despite the different primary sequences of the hairpins, the molecular logic and precise cross-regulation of Microprocessor components is conserved between *Drosophila* and mammals.

### Synaptic function is sensitive to global reduction of miRNA levels in photoreceptor neurons

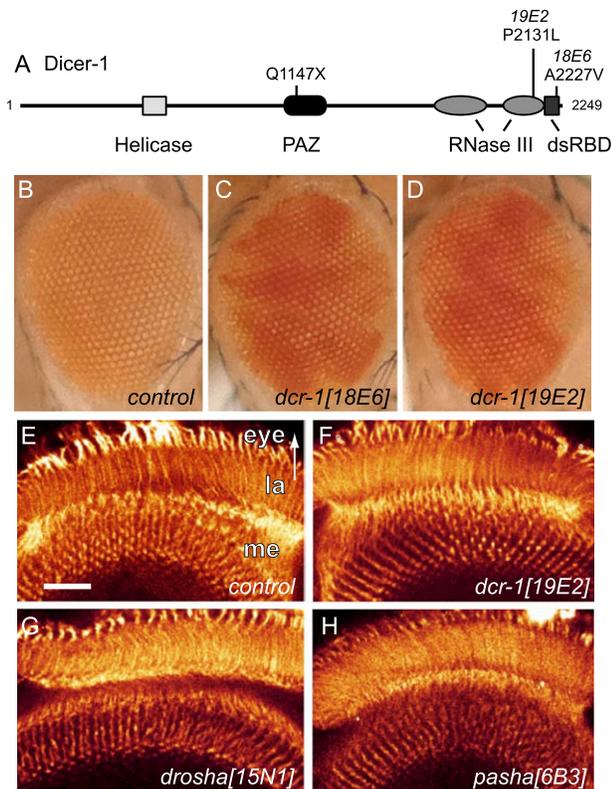
In *Drosophila*, as in other organisms, loss of core components of the miRNA pathway causes a plethora of developmental consequences (Lee et al. 2004; Hatfield et al. 2005; Jin and Xie 2007; Park et al. 2007; Martin et al. 2009). However, it is difficult even with conditional knockout techniques to bypass developmental effects. Our hypomorphic mutants of core

miRNA pathway components provided an opportunity to assess the relative sensitivity of development, function, and maintenance on miRNA levels in a specialized cell type. In particular, the observation that the misregulation of certain miRNA targets did not strictly correlate with overall status of

the miRNA pathway suggested that it might be possible to discern specific cellular settings that are especially sensitive to miRNA levels.

In addition to our weak *drosha*[15N1] and *pasha*[6B3] alleles, our screen isolated novel alleles of *dcr-1*, *dcr-1*[18E6], and *dcr-1*[19E2], which have missense mutations in the double-stranded RNA binding domain and the second RNase III domain, respectively (Fig. 5A). These hypomorphic mutations exhibited w-miR derepression and adult eye clonal morphology reminiscent of our previously described *pasha*[6B3] and *drosha*[15N1] hypomorphic alleles (Figs. 2E,N, 5B–D). They yielded morphologically normal, pharate adult lethal animals with some escapers *in trans* with either a *dcr-1*-null mutation, *dcr-1*[Q1147X], or a deficiency that removes the *dcr-1* locus, *Df(3R)ED6906* (Lee et al. 2004; Ryder et al. 2007). Together with the hypomorphic *pasha*[6B3] and *drosha*[15N1] alleles, we used these reagents to assess post-developmental roles for the miRNA pathway in the eye.

Since external morphology does not reliably report on possible defects of internal structures, we first performed



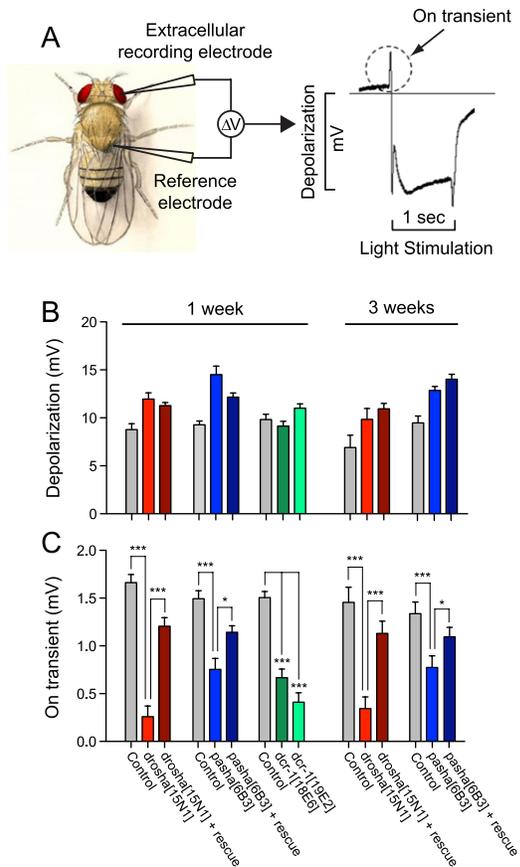
**FIGURE 5.** Hypomorphic miRNA pathway mutants exhibit normal eye development. (A) Schematic representation of Dcr-1 protein with positions of missense mutations *dcr-1*[18E6] and *dcr-1*[19E2] and nonsense mutation *dcr-1*[Q1147X] (Lee et al. 2004) indicated. (B) Control *w-miR* eye. Eyes bearing clones of *dcr-1*[18E6] (C) or *dcr-1*[19E2] (D) in the background of *w-miR* exhibit derepression of the white reporter, seen as darker red pigmentation, but otherwise lack roughness that might indicate aberrant development. (E–H) In comparison to wild type, hypomorphic mutants of *dcr-1*, *drosha*, and *pasha* all exhibit normal patterns of photoreceptor projections to the central brain.

additional analysis on animals bearing whole eye clones of hypomorphic miRNA biogenesis mutants. These tests indicated normal specification of photoreceptor and support cells. Moreover, the mutant photoreceptors retained normal targeting properties and established layer-specific terminals in the optic neuropils lamina and medulla that are indistinguishable from wild type (Fig. 5E–H). These findings demonstrated that development and wiring of the eye was not substantially affected by a mild reduction in miRNA biogenesis.

Photoreceptor neuron function and maintenance (or degeneration) is easily assessed through electroretinogram recordings (Williamson et al. 2010). Electroretinograms (ERGs) are extracellular recordings of the photoreceptor response to a light stimulus that exhibit two main features: The depolarization amplitude, which is reduced when phototransduction or cell health is defective or degrades over time, and the “on” transient, a post-synaptic response that is lost when synaptic transmission is blocked (Fig. 6A). Hence, ERGs provide a measure for both neuronal degeneration and function over time.

We performed ERGs on whole eye clones of our miRNA pathway hypomorphic mutants using the *ey3.5FLP* system, which selectively renders the pre-synaptic photoreceptor neurons mutant (Fig. 6B; Chotard et al. 2005; Mehta et al. 2005). Whole eye clones of *pasha*[6B3] showed no substantial difference in the amplitude of depolarization compared with control eyes, indicating that this mutation does not affect the photoreceptor response to stimulation. However, *pasha*[6B3] ERGs exhibited significantly decreased “on” transients. This defect is significantly rescued by including a single copy of a *pasha* genomic rescue construct in the *pasha*[6B3] background. We saw a stronger reduction of “on” transient amplitude in *drosha*[15N1] whole eye clones, which we could also rescue to near wild-type levels with a single copy of a *drosha* genomic rescue construct. In addition, our two independent hypomorphic *dcr-1* alleles displayed significant reduction in “on” transient amplitude with no significant effect on depolarization (Fig. 6B). The synaptic transmission defects cannot be attributed to post-synaptic neurons because only the pre-synaptic photoreceptors were mutant in these experiments. Taken together, our findings reveal a specific loss of neurotransmission in hypomorphic alleles for *drosha*, *pasha*, and *dcr-1* that all mildly reduce miRNA levels without affecting development. These data suggest that synaptic transmission is more sensitive to miRNA levels than development of photoreceptor neurons.

It was conceivable that a gradual loss of synaptic transmission precedes neuronal degeneration in the presence of reduced miRNA pathway activity. To test this notion, we compared the depolarization and “on” transients of *drosha*[15N1] and *pasha*[6B3] in 1-wk and 3-wk aged flies. As shown in Figure 6B,C, the recordings were virtually identical. We conclude that neither the healthy depolarization upon light stimulation nor the synaptic transmission



**FIGURE 6.** Hypomorphic alleles of miRNA pathway components have specific synaptic transmission defects. (A) Illustration of the ERG procedure and description of measurements. Electroretinograms (ERGs) were performed on animals with whole eye clones of the indicated mutant alleles (see also Materials and Methods). (B) Depolarization and (C) “on” transient amplitude measurements of indicated genotypes. Whole eye clones of *drosha*[15N1] and *pasha*[6B3] result in significant loss of “on” transient amplitude that is rescued by a single copy of a genomic rescue transgene. Two independent alleles of *dcr-1* also exhibited a significant loss of “on” transient amplitude. In all cases, depolarization of the photoreceptor cell bodies was not significantly different from controls. ERG analysis in 3-wk-old flies revealed no change in “on” transient amplitude or depolarization in any genotypes examined, demonstrating an absence of large-scale photoreceptor degradation or death. Values represent the mean depolarization or “on” transient amplitude of >10 flies per genotype  $\pm$  standard error of the mean. \*\*\* $P < 0.01$ , \* $P < 0.05$ .

defect deteriorates over time. These findings suggest that the reduced neurotransmission in the Microprocessor hypomorphs is not secondary to neuronal maintenance defects and, reciprocally, that miRNA function is acutely required for synaptic function.

## DISCUSSION

### An efficient in vivo genetic screen for miRNA pathway components

In this study, we describe a forward genetic screen for factors involved in miRNA biogenesis or function and

validate its utility by characterizing a series of core miRNA pathway mutants. We used these to investigate Microprocessor cross-regulation in vivo, as well as to study post-developmental roles of miRNAs in neural function. In particular, we provide the first loss-of-function analysis of *Drosophila drosha*. As expected, we observed a strong block in canonical miRNA biogenesis in the *drosha*-null mutant, which accumulated primary miRNA transcripts and was depleted of mature miRNAs, similar to *pasha* mutants (Martin et al. 2009). In addition, we found mirtron biogenesis to be unaffected by *drosha* mutation, providing direct evidence that these splicing-derived miRNAs are completely independent of the Drosha nuclease. Animals null for *drosha* and *pasha* are generally similar with respect to all phenotypes examined, supporting the obligate nature of these core protein partners within the Microprocessor complex.

Our screen generated allelic series for the key Microprocessor components *drosha* and *pasha*. These allowed us for the first time to assess the sensitivities of development versus function in cells with mildly reduced miRNA levels. Compared with full knockouts that give only null states, forward genetic screening can yield allelic series of varying phenotypic severity, which can uncover interesting aspects of pathway and gene function. We note that our hypomorphic *drosha* and *pasha* mutants have very different effects in different assays, as highlighted by the differences in derepression of both an endogenous miRNA target and a transgenic sensor for miRNA activity between our weak *drosha* and *pasha* alleles (Fig. 2). This is the case despite the fact that these mutants have similar effects on total mature miRNA levels as measured by Northern blot (Fig. 3B). The reason for this phenotypic discrepancy is not yet clear but suggests the possibilities that miRNAs are differentially sensitive to availability of the core biogenesis machinery or that specific biological processes are especially sensitive to optimal miRNA biogenesis. Indeed, we have provided electrophysiological evidence of the latter, and both of these hypotheses represent compelling future directions for analysis of these and other mutants that may emerge from genetic screening.

The theme of miRNA pathway autoregulation has emerged at multiple levels in animals (Forman et al. 2008; Han et al. 2009; Kadener et al. 2009; Triboulet et al. 2009; Martello et al. 2010; Bennasser et al. 2011) and in plants (Xie et al. 2003; Vaucheret et al. 2004; Rajagopalan et al. 2006; Vaucheret et al. 2006). Our in vivo demonstration of the reciprocal regulation of the core Microprocessor components reveals that this mechanism is conserved and occurs within the intact animal. The role of Pasha within the Microprocessor to position Drosha catalytic centers is clearly of crucial importance to miRNA biogenesis. Perhaps the instability of Drosha in the absence of Pasha is a biological safeguard to prevent inappropriate cleavage of transcripts by Drosha in the absence of Pasha. Likewise, the capacity of Drosha to cleave *pasha* transcripts may also limit Drosha

levels by restricting the amount of Pasha for it to associate with.

Our studies to date focused on mutants of known miRNA pathway components. This has been a productive effort, as indicated by our first reported *Drosophila* alleles of *drossha*, the first allelic series of *pasha* in any organism, and novel alleles of *dicer-1*. Our screening recovered other suppressor mutations that do not map to known pathway components, as well as enhancer mutations that are dependent on the *w-miR* trigger. We anticipate that the cloning and characterization of these mutations may provide additional insights into the miRNA biogenesis machinery or the mechanism of miRNA-mediated silencing.

### Hypomorphic alleles reveal a synaptic transmission defect caused by mild reduction of miRNA pathway activity

miRNAs in whole organisms have to date mostly been studied for their roles in development. This is at least in part due to the early developmental defects that potentially mask later functional defects. Yet, miRNAs have long been viewed as a potentially key component of neural function and fine-tuning due to their regulatory potential. One tantalizing property of miRNAs is their ability to regulate gene expression locally. In neurons, where a synapse may be a great distance from the nucleus, this could provide a means for rapid post-transcriptional regulation of targets. How this may be regulated in a signal-dependent manner is a topic of ongoing study (Ashraf et al. 2006; Krol et al. 2010). In this study, we present novel hypomorphic mutants that mildly affect miRNA levels and cause synapse function defects without affecting development.

The requirement of neurons for precise miRNA activity is emphasized by the specific synaptic transmission defects in our hypomorphic miRNA pathway mutants. In weak alleles of either *pasha* or *drossha*, we observed only a mild reduction in miRNA biogenesis. While these animals quickly succumb just before or after adult eclosion, they exhibit grossly normal development of all external structures. Using whole eye clone analysis, we documented normal specification and projection of mutant photoreceptors, but substantial synaptic transmission defects that were very similar in *drossha*, *pasha*, and *dicer-1* mutants. The lack of deterioration of this phenotype with age both argues for a specific synaptic transmission defect and also that the reduction in miRNA levels in these photoreceptor neurons does not affect their viability or general health.

The commonality of these phenotypes among the different mutants indicates underlying defective biogenesis of one or more canonical miRNAs, as opposed to mirtrons or other noncanonical species. It is conceivable that the synaptic transmission defect is caused by the cumulative effect of mildly reducing all photoreceptor miRNAs. However, the specificity of this phenotype and its critical dependence on

optimal miRNA pathway function may imply that there is some aspect of synaptic transmission machinery that is especially sensitive to a more limited set of miRNAs. As the number of mutant strains for *Drosophila* miRNA loci steadily increases, it will be productive to screen them using ERGs on mutant eyes. An alternative approach may be to test miRNA sponges (Loya et al. 2009) expressed pre-synaptically, perhaps in a candidate screen of head-expressed miRNAs (Ruby et al. 2007b).

Since loss of single alleles is typically well tolerated at the organismal level, dose-sensitive loci are of particular relevance to human disease. DGCR8 is one of about 20 genes within the 22q11.2 locus deleted in patients with diGeorge syndrome, for which heterozygosity results in cognitive and behavioral impairments in humans. In a mouse model of diGeorge syndrome bearing the 22q11.2 microdeletion, heterozygosity of *dgcr8* contributes to a reduction in brain miRNAs (Stark et al. 2008). More recently, specific heterozygosity of *dgcr8* was shown to cause subtle but detectable defects in neural developmental and physiology, providing more direct evidence of *dgcr8* haploinsufficiency (Fenelon et al. 2011). In the *Drosophila* system, cells heterozygous for *pasha*[*KO*] have less Pasha protein than homozygous wild-type cells (Fig. 4H). Accordingly, the levels of the GFP-ban sensor indicated that it was repressed more strongly in homozygous wild-type cells than in neighboring *pasha*[*KO*] heterozygous cells (Fig. 2I). Therefore, despite the operation of feedback mechanisms that regulate Microprocessor levels, *pasha* exhibits functional haploinsufficiency. Altogether, our studies reaffirm that subtle alterations in miRNA biogenesis can lead to detectable organismal phenotypes, helping to explain the lengths to which animal evolution has gone to maintain Microprocessor homeostasis. Reciprocally, our studies define *Drosophila* as a suitable system for future exploration of the unique sensitivity of neuronal function to miRNA activity.

## MATERIALS AND METHODS

### Molecular biology and biochemistry

#### Generation of *w-miR*

pGUS-DsRed was generated by cloning the DsRed.T4 CDS from Red-H-Pelican (Barolo et al. 2004) as an NcoI(blunt)/NotI fragment into pGUS (Brodsky et al. 2000) cut with EcoRI(blunt)/NotI. We used the strategy of Hay and colleagues to reprogram the *mir-6-1* backbone (Chen et al. 2007) to produce an miR targeted against the *white* 3' UTR. The partially complementary primers mir6.1 wmiRNA1 For: GGCAGCTTACTTAACTTAATCACAGCCTTT AATGTTTACCATCGTCGGCACTGTCTATAAGTTAATATACCA TATC and mir6.1 wmiRNA1 Rev: AATAATGATGTTAGGCACT TTAGGTACTTACCATCGTCAATACTGTTTATAGATATGGTAT ATTAAGTTATAGA were converted into a dsDNA template by PCR and used as template for a second round of PCR using primers mir6.1 5' NotI: gatcgcgccgcgTTTTAAAGTCCACAATC ATCAAGGAAAATGAAAGTCAAAGTTGGCAGCTTACTTAAAC

TTA and mir6.1 3' XbaI: gatctctagaAAAACGGCATGGTTATTCG TGTGCCAAAAAAAAAAAAAAAAATTAATAATGATGTTAGGCAC. This product was digested with NotI and XbaI and cloned into pGUS-DsRed. Transgenic animals were generated by standard *P*-element-mediated transformation (Bestgene). Lines with consistent pale-orange eye color on chromosomes II and III were recombined with eyFLP transgenes on these respective chromosomes. The presence of the eyFLP transgene was confirmed by PCR to the *flp* coding region.

#### *Droscha* rescue construct

The *droscha* genomic locus from the isogenized FRT42 strain we used for mutagenesis was amplified in two parts using Droscha F FseI: gatcgccggccAACTGATCCGCTTCCTGGCG and Droscha mid R: TAATGCCGGTGGGATAGAAAGC for the 5' half and Droscha mid F: GATGGCCAACATGTCCAAGCC and Droscha R NotI: gatcgccggccCAACGTCTCCTGAGCTCGCC for the 3' half. The overlapping region between these PCR products contains a unique PciI restriction site. The 5' half was digested with FseI/PciI and the 3' half with PciI/NotI. These fragments were cloned into pBDP (Pfeiffer et al. 2008) cut with FseI/NotI. Transgenic animals were made by targeted transgenesis into the attP2 site on chromosome 3L.

#### *Pasha* rescue transgene lacking 5'-UTR hairpins

We cloned our previously described *pasha* genomic rescue construct (Martin et al. 2009) into pBDP, and the sequences comprising the 5'-UTR hairpins were replaced with unstructured sequences from *GFP*. For a detailed description of this construction, see Supplemental Figure 1.

#### *white* 3'-UTR sensors

The annotated *white* 3' UTR and ~150 bases of downstream sequence was amplified with w 3' UTR Not For: gatcgccggccCGG ACATATATCCGAAATAAC and w 3' UTR Xho Rev: gatcctc gagCTATAGGTCATATCTTGTTTT. The PCR product was digested with NotI and XhoI and cloned into both tub-GFP (Stark et al. 2003) and modified psiCHECK2 (Okamura et al. 2007). Luciferase sensor assays in S2 cells were performed as previously described (Okamura et al. 2007).

#### Sequencing of mutant alleles

Following identification of mutant alleles by complementation, alleles of interest were placed *in trans* with *pasha*[*KO*] (*pasha* alleles) or *Df*(2R)*exel6055* (*droscha* alleles). Larvae hemizygous for the locus of interest were selected, and DNA was extracted and amplified by PCR using the same oligos used to generate genomic rescue constructs. The molecular lesions associated with each allele were determined by standard sequencing and comparison with the sequence of the nonmutagenized starting strain.

#### Analysis of miRNA/mirtron processing

qRT-PCR analyses were performed using previously described oligo sequences and protocols (Martin et al. 2009). For Northern analysis, we used TRIzol to isolate total RNA from wild-type larvae (Canton S) or larvae *trans*-heterozygous for the indicated mutant allele and

*pasha*[*KO*] (for *pasha* alleles) or *Df*(2R)*exel6055* (for *droscha* alleles). Small RNA Northern blots were performed using polyacrylamide gels and LNA probes (Exiqon) as described (Okamura et al. 2007).

#### *Drosophila* genetic screen

We mutagenized flies by collecting about 50–70 males and starved them by placing them in a dry empty vial with Whatman paper for ~8 h. Flies were then fed overnight with a solution of 25 mM EMS in 20% sucrose injected onto the Whatman paper. The males were then given 8 h to recover on normal food before being mated with females in a freshly yeasted bottle. Typically, five vials of males were mutagenized in each round, and mated females were allowed to lay on three consecutive sets of bottles for 2 d each. The adult flies were then discarded.

The screen for chromosome 3R was performed as follows. Virgin females of the genotype *w*; *GMR-w-miR*, *eyFLP*; *iso FRT82B* were crossed with mutagenized males of the genotype *w*; *Sco/CyO*; *isoFRT82B*. F<sub>1</sub> progeny with eye color mosaicism were crossed again with *w*; *GMR-w-miR*, *eyFLP*; *isoFRT82B* to check for germline transmission of the mutation. Progeny displaying mosaicism were crossed to *w*; *Sco/CyO*; *MKRS/TM6B*. As the mutagenized chromosome is superficially impossible to distinguish from the nonmutagenized chromosome, individual males (about 10) were then crossed to *w*; *Sco/CyO*; *MKRS/TM6B*, and stocks were generated that were *w*; *Sco/CyO*; *FRT82B*, *mutant?/TM6B*. Half of these stocks are expected to have the mutagenized *FRT82B* chromosome. One stock per original mutant that could reproduce the original phenotype when crossed to *w*; *GMR-w-miR*, *eyFLP*; *iso FRT82B* was kept. The screen on chromosome 2R was carried out with the same strategy as described for 3R, with the exception that an isogenized FRT42D chromosome and a *GMR-w-miR*, *eyFLP* chromosome III were used instead. Images of eye clones in adult animals presented are of the genotype *w*; *GMR-w-miR*, *eyFLP/Sco* or *CyO*; *FRT82* [*X*]/*FRT82*, where [*X*] denotes the appropriate *pasha* or *dcr-1* allele or *w*; *FRT42* [*Y*]/*FRT42*; *GMR-w-miR*, *eyFLP/MKRS* or *TM6B*, where [*Y*] denotes the appropriate *droscha* allele.

#### Clonal analysis and immunostaining

We analyzed the functional properties of mutants isolated in this screen, as well as *FRT82B pasha*[*KO*] (Martin et al. 2009) and *FRT82B dcr-1*[*Q1147X*] (Lee et al. 2004), by introducing them into backgrounds carrying *hs-FLP* and the miRNA sensor transgenes *tub-GFP-bantam* (Brennecke et al. 2003) or *tub-GFP-dLMO 3' UTR* (Bejarano et al. 2010). Chromosome 3R clones were generated in the genotype *hs-FLP*; *GFP-ban/+*; *FRT82B* [*X*]/*FRT82B*, *arm-lacZ* (Fig. 2) or *hs-FLP*; *FRT82B* [*X*]/*FRT82B*, *arm-lacZ* (Fig. 5), where [*X*] denotes the relevant mutant allele. Chromosome 2R clones were generated in the genotype *hs-FLP*; *FRT42D* [*Y*]/*FRT42D*, *arm-lacZ*; *GFP-dLMO/+* (Fig. 2) or *hs-FLP*; *FRT42D* [*Y*]/*FRT42D*, *arm-lacZ* (Fig. 5), where [*Y*] denotes the appropriate mutant allele. For Figure 5G, the genotype analyzed was *hs-FLP*; *FRT42D*, *droscha*[*21K11*]/*FRT42D*, *arm-lacZ*; *attP2>pBDP pasha*[ $\Delta$ H $P$ ] rescue, *pasha*[*KO*].

Immunostaining was performed as previously described (Lai and Rubin 2001). Antibodies used were mouse anti- $\beta$ -galactosidase (1/50) and rat anti-Elav (1/50) from the Developmental Studies Hybridoma Bank, Chicken anti-GFP (1/1000, Abcam),

rabbit anti-Mei-P26 (1/1000) (Liu et al. 2009), rabbit anti-Pasha (1/1000), and rabbit anti-Drosha (1/1000) (Denli et al. 2004). We used appropriate secondary antibodies conjugated to Alexa 488, 568, and 647 (Molecular Probes).

## Electrophysiology

Electroretinograms (ERGs) were performed as described in Fabian-Fine et al. (2003) with the following modifications: Flies were fixed using Elmer's nontoxic Glue-All. We used 2 M NaCl in the recording and reference electrodes. Recordings were performed using a Warner IE-210 amplifier and Clampex 10.1 software by Axon Instruments. A post-recording filter was also provided by the Clampex software. Light stimulus was provided in 1-sec pulses by a computer-controlled white LED system (Schott MC1500).

The following genotypes were used:

*ey3.5FLP; FRT42D, CL/CyO (FRT42D Ctrl)*  
*ey3.5FLP; FRT42D, CL/FRT42D, drosha[15N1] (FRT42 drosha)*  
*ey3.5FLP; FRT42D, CL/FRT42D, drosha[15N1]; attP2>BDP drosha/+ (FRT42 drosha rescue)*  
*ey3.5FLP; ; FRT82B, CL/TM3, Sb (FRT82 Ctrl)*  
*ey3.5FLP; ; FRT82B, CL/FRT82B, pasha[6B3] (FRT82 pasha 6B3)*  
*ey3.5FLP; ; FRT82B, CL/attP2>BDP pasha-FlagHA, FRT82B, pasha[6B3] (FRT82 pasha 6B3 rescue)*  
*ey3.5FLP; ; FRT82B, CL/FRT82B, dcr-1[18E6]*  
*ey3.5FLP; ; FRT82B, CL/FRT82B, dcr-1[19E2]*

## SUPPLEMENTAL MATERIAL

Supplemental material is available for this article.

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## REFERENCES

Ashraf SI, McLoon AL, Scarsic SM, Kunes S. 2006. Synaptic protein synthesis associated with memory is regulated by the RISC pathway in *Drosophila*. *Cell* **124**: 191–205.  
 Babiarz JE, Ruby JG, Wang Y, Bartel DP, Blelloch R. 2008. Mouse ES cells express endogenous shRNAs, siRNAs, and other Microprocessor-independent, Dicer-dependent small RNAs. *Genes Dev* **22**: 2773–2785.

Barolo S, Castro B, Posakony JW. 2004. New *Drosophila* transgenic reporters: insulated P-element vectors expressing fast-maturing RFP. *BioTechniques* **36**: 436–442.  
 Bartel DP. 2009. MicroRNAs: Target recognition and regulatory functions. *Cell* **136**: 215–233.  
 Behm-Ansmant I, Rehwinkel J, Doerks T, Stark A, Bork P, Izaurralde E. 2006. mRNA degradation by miRNAs and GW182 requires both CCR4:NOT deadenylase and DCP1:DCP2 decapping complexes. *Genes Dev* **20**: 1885–1898.  
 Bejarano F, Smibert P, Lai EC. 2010. miR-9a prevents apoptosis during wing development by repressing *Drosophila* LIM-only. *Dev Biol* **338**: 63–73.  
 Bannasser Y, Chable-Bessia C, Triboulet R, Gibbings D, Gwizdek C, Dargemont C, Kremer EJ, Voinnet O, Benkirane M. 2011. Competition for XPO5 binding between Dicer mRNA, pre-miRNA and viral RNA regulates human Dicer levels. *Nat Struct Mol Biol* **18**: 323–327.  
 Berdnik D, Fan AP, Potter CJ, Luo L. 2008. MicroRNA processing pathway regulates olfactory neuron morphogenesis. *Curr Biol* **18**: 1754–1759.  
 Bohnsack MT, Czaplinski K, Gorlich D. 2004. Exportin 5 is a RanGTP-dependent dsRNA-binding protein that mediates nuclear export of pre-miRNAs. *RNA* **10**: 185–191.  
 Brenneke J, Hipfner DR, Stark A, Russell RB, Cohen SM. 2003. *bantam* encodes a developmentally regulated microRNA that controls cell proliferation and regulates the proapoptotic gene *hid* in *Drosophila*. *Cell* **113**: 25–36.  
 Brodsky MH, Nordstrom W, Tsang G, Kwan E, Rubin GM, Abrams JM. 2000. *Drosophila* p53 binds a damage response element at the *reaper* locus. *Cell* **101**: 103–113.  
 Cheloufi S, Dos Santos CO, Chong MM, Hannon GJ. 2010. A dicer-independent miRNA biogenesis pathway that requires Ago catalysis. *Nature* **465**: 584–589.  
 Chen CH, Huang H, Ward CM, Su JT, Schaeffer LV, Guo M, Hay BA. 2007. A synthetic maternal-effect selfish genetic element drives population replacement in *Drosophila*. *Science* **316**: 597–600.  
 Chong MM, Rasmussen JP, Rudensky AY, Littman DR. 2008. The RNaseIII enzyme Drosha is critical in T cells for preventing lethal inflammatory disease. *J Exp Med* **205**: 2005–2017.  
 Chotard C, Leung W, Salecker I. 2005. *glial cells missing* and *gcm2* cell autonomously regulate both glial and neuronal development in the visual system of *Drosophila*. *Neuron* **48**: 237–251.  
 Cifuentes D, Xue H, Taylor DW, Patnode H, Mishima Y, Cheloufi S, Ma E, Mane S, Hannon GJ, Lawson N, et al. 2010. A novel miRNA processing pathway independent of Dicer requires Argonaute2 catalytic activity. *Science* **328**: 1694–1698.  
 Czech B, Zhou R, Erlich Y, Brenneke J, Binari R, Villalta C, Gordon A, Perrimon N, Hannon GJ. 2009. Hierarchical rules for Argonaute loading in *Drosophila*. *Mol Cell* **36**: 445–456.  
 Denli A, Tops B, Plasterk R, Ketting R, Hannon G. 2004. Processing of primary microRNAs by the Microprocessor complex. *Nature* **432**: 231–235.  
 Eulalio A, Rehwinkel J, Stricker M, Huntzinger E, Yang SF, Doerks T, Dorner S, Bork P, Boutros M, Izaurralde E. 2007. Target-specific requirements for enhancers of decapping in miRNA-mediated gene silencing. *Genes Dev* **21**: 2558–2570.  
 Eulalio A, Huntzinger E, Izaurralde E. 2008. GW182 interaction with Argonaute is essential for miRNA-mediated translational repression and mRNA decay. *Nat Struct Mol Biol* **15**: 346–353.  
 Fabian-Fine R, Verstreken P, Hiesinger PR, Horne JA, Kostyleva R, Zhou Y, Bellen HJ, Meinertzhagen IA. 2003. Endophilin promotes a late step in endocytosis at glial invaginations in *Drosophila* photoreceptor terminals. *J Neurosci* **23**: 10732–10744.  
 F enelon K, Mukai J, Xu B, Hsu PK, Drew LJ, Karayiorgou M, Fischbach GD, Macdermott AB, Gogos JA. 2011. Deficiency of *Dgcr8*, a gene disrupted by the 22q11.2 microdeletion, results in altered short-term plasticity in the prefrontal cortex. *Proc Natl Acad Sci* **108**: 4447–4452.

- Flynt AS, Lai EC. 2008. Biological principles of microRNA-mediated regulation: shared themes amid diversity. *Nat Rev Genet* **9**: 831–842.
- Flynt AS, Chung WJ, Greimann JC, Lima CD, Lai EC. 2010. microRNA biogenesis via splicing and exosome-mediated trimming in *Drosophila*. *Mol Cell* **38**: 900–907.
- Forman JJ, Legesse-Miller A, Collier HA. 2008. A search for conserved sequences in coding regions reveals that the *let-7* microRNA targets Dicer within its coding sequence. *Proc Natl Acad Sci* **105**: 14879–14884.
- Förstemann K, Tomari Y, Du T, Vagin V, Denli A, Bratu D, Klattenhoff C, Theurkauf W, Zamore P. 2005. Normal microRNA maturation and germ-line stem cell maintenance requires Loquacious, a double-stranded RNA-binding domain protein. *PLoS Biol* **3**: e236. doi: 10.1371/journal.pbio.0030236.
- Förstemann K, Horwich MD, Wee L, Tomari Y, Zamore PD. 2007. *Drosophila* microRNAs are sorted into functionally distinct argonaute complexes after production by Dicer-1. *Cell* **130**: 287–297.
- Fukuda T, Yamagata K, Fujiyama S, Matsumoto T, Koshida I, Yoshimura K, Mihara M, Naitou M, Endoh H, Nakamura T, et al. 2007. DEAD-box RNA helicase subunits of the Drosha complex are required for processing of rRNA and a subset of microRNAs. *Nat Cell Biol* **9**: 604–611.
- Ghildiyal M, Xu J, Seitz H, Weng Z, Zamore PD. 2010. Sorting of *Drosophila* small silencing RNAs partitions microRNA\* strands into the RNA interference pathway. *RNA* **16**: 43–56.
- Gregory RI, Yan KP, Amuthan G, Chendrimada T, Doratotaj B, Cooch N, Shiekhattar R. 2004. The Microprocessor complex mediates the genesis of microRNAs. *Nature* **432**: 235–240.
- Han J, Lee Y, Yeom KH, Kim YK, Jin H, Kim VN. 2004. The Drosha-DGCR8 complex in primary microRNA processing. *Genes Dev* **18**: 3016–3027.
- Han J, Lee Y, Yeom KH, Nam JW, Heo I, Rhee JK, Sohn SY, Cho Y, Zhang BT, Kim VN. 2006. Molecular basis for the recognition of primary microRNAs by the Drosha-DGCR8 complex. *Cell* **125**: 887–901.
- Han J, Pedersen JS, Kwon SC, Belair CD, Kim YK, Yeom KH, Yang WY, Haussler D, Blelloch R, Kim VN. 2009. Posttranscriptional crossregulation between Drosha and DGCR8. *Cell* **136**: 75–84.
- Hatfield SD, Shcherbata HR, Fischer KA, Nakahara K, Carthew RW, Ruohola-Baker H. 2005. Stem cell division is regulated by the microRNA pathway. *Nature* **435**: 974–978.
- Herranz H, Hong X, Perez L, Ferreira A, Olivieri D, Cohen SM, Milan M. 2010. The miRNA machinery targets Mei-P26 and regulates Myc protein levels in the *Drosophila* wing. *EMBO J* **29**: 1688–1698.
- Jin Z, Xie T. 2007. Dcr-1 maintains *Drosophila* ovarian stem cells. *Curr Biol* **17**: 539–544.
- Kadener S, Rodriguez J, Abruzzi KC, Khodor YL, Sugino K, Marr MT II, Nelson S, Rosbash M. 2009. Genome-wide identification of targets of the drosha-pasha/DGCR8 complex. *RNA* **15**: 537–545.
- Kim K, Lee YS, Carthew RW. 2007. Conversion of pre-RISC to holo-RISC by Ago2 during assembly of RNAi complexes. *RNA* **13**: 22–29.
- Kim VN, Han J, Siomi MC. 2009. Biogenesis of small RNAs in animals. *Nat Rev Mol Cell Biol* **10**: 126–139.
- Krol J, Busskamp V, Markiewicz I, Stadler MB, Ribi S, Richter J, Duebel J, Bicker S, Fehling HJ, Schubeler D, et al. 2010. Characterizing light-regulated retinal microRNAs reveals rapid turnover as a common property of neuronal microRNAs. *Cell* **141**: 618–631.
- Lai EC. 2002. microRNAs are complementary to 3' UTR sequence motifs that mediate negative post-transcriptional regulation. *Nat Genet* **30**: 363–364.
- Lai EC, Posakony JW. 1997. The Bearded box, a novel 3' UTR sequence motif, mediates negative post-transcriptional regulation of *Bearded* and *Enhancer of split* Complex gene expression. *Development* **124**: 4847–4856.
- Lai EC, Rubin GM. 2001. *neuritized* functions cell-autonomously to regulate a subset of Notch-dependent processes during adult *Drosophila* development. *Dev Biol* **231**: 217–233.
- Lai EC, Burks C, Posakony JW. 1998. The K box, a conserved 3' UTR sequence motif, negatively regulates accumulation of *Enhancer of split* complex transcripts. *Development* **125**: 4077–4088.
- Landthaler M, Yalcin A, Tuschl T. 2004. The human DiGeorge syndrome critical region gene 8 and its *D. melanogaster* homolog are required for miRNA biogenesis. *Curr Biol* **14**: 2162–2167.
- Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, Lee J, Provost P, Radmark O, Kim S, et al. 2003. The nuclear RNase III Drosha initiates microRNA processing. *Nature* **425**: 415–419.
- Lee YS, Nakahara K, Pham JW, Kim K, He Z, Sontheimer EJ, Carthew RW. 2004. Distinct roles for *Drosophila* Dicer-1 and Dicer-2 in the siRNA/miRNA silencing pathways. *Cell* **117**: 69–81.
- Lee YS, Pressman S, Andress AP, Kim K, White JL, Cassidy JJ, Li X, Lubell K, Lim DH, Cho IS, et al. 2009. Silencing by small RNAs is linked to endosomal trafficking. *Nat Cell Biol* **11**: 1150–1156.
- Lim H, Kim J, Kim S, Carthew RW, Lee YS. 2008. Functional analysis of *dicer-2* missense mutations in the siRNA pathway of *Drosophila*. *Biochem Biophys Res Commun* **371**: 525–530.
- Liu J, Rivas FV, Wohlschlegel J, Yates JR III, Parker R, Hannon GJ. 2005. A role for the P-body component GW182 in microRNA function. *Nat Cell Biol* **7**: 1161–1166.
- Liu N, Han H, Lasko P. 2009. Vasa promotes *Drosophila* germline stem cell differentiation by activating *mei-P26* translation by directly interacting with a (U)-rich motif in its 3' UTR. *Genes Dev* **23**: 2742–2752.
- Loya CM, Lu CS, Van Vactor D, Fulga TA. 2009. Transgenic microRNA inhibition with spatiotemporal specificity in intact organisms. *Nat Methods* **6**: 897–903.
- Lund E, Guttinger S, Calado A, Dahlberg J, Kutay U. 2004. Nuclear export of microRNA precursors. *Science* **303**: 95–98.
- Marques JT, Kim K, Wu PH, Alleyne TM, Jafari N, Carthew RW. 2010. *Loqs* and *R2D2* act sequentially in the siRNA pathway in *Drosophila*. *Nat Struct Mol Biol* **17**: 24–30.
- Martello G, Rosato A, Ferrari F, Manfrin A, Cordenonsi M, Dupont S, Enzo E, Guzzardo V, Rondina M, Spruce T, et al. 2010. A microRNA targeting dicer for metastasis control. *Cell* **141**: 1195–1207.
- Martin R, Smibert P, Yalcin A, Tyler DM, Schaefer U, Tuschl T, Lai EC. 2009. A *Drosophila pasha* mutant distinguishes the canonical miRNA and mirtron pathways. *Mol Cell Biol* **29**: 861–870.
- Mehta SQ, Hiesinger PR, Beronja S, Zhai RG, Schulze KL, Verstreken P, Cao Y, Zhou Y, Tepass U, Crair MC, et al. 2005. Mutations in *Drosophila sec15* reveal a function in neuronal targeting for a subset of exocyst components. *Neuron* **46**: 219–232.
- Okamura K, Hagen JW, Duan H, Tyler DM, Lai EC. 2007. The mirtron pathway generates microRNA-class regulatory RNAs in *Drosophila*. *Cell* **130**: 89–100.
- Okamura K, Liu N, Lai EC. 2009. Distinct mechanisms for microRNA strand selection by *Drosophila* Argonautes. *Mol Cell* **36**: 431–444.
- Park JK, Liu X, Strauss TJ, McKearin DM, Liu Q. 2007. The miRNA pathway intrinsically controls self-renewal of *Drosophila* germline stem cells. *Curr Biol* **17**: 533–538.
- Parry DH, Xu J, Ruvkun G. 2007. A whole-genome RNAi screen for *C. elegans* miRNA pathway genes. *Curr Biol* **17**: 2013–2022.
- Pfeiffer BD, Jenett A, Hammonds AS, Ngo TT, Misra S, Murphy C, Scully A, Carlson JW, Wan KH, Laverty TR, et al. 2008. Tools for neuroanatomy and neurogenetics in *Drosophila*. *Proc Natl Acad Sci* **105**: 9715–9720.
- Rajagopalan R, Vaucheret H, Trejo J, Bartel DP. 2006. A diverse and evolutionarily fluid set of microRNAs in *Arabidopsis thaliana*. *Genes Dev* **20**: 3407–3425.
- Rehwinkel J, Behm-Ansmant I, Gatfield D, Izaurralde E. 2005. A crucial role for GW182 and the DCP1:DCP2 decapping complex in miRNA-mediated gene silencing. *RNA* **11**: 1640–1647.
- Ruby JG, Jan CH, Bartel DP. 2007a. Intronic microRNA precursors that bypass Drosha processing. *Nature* **448**: 83–86.
- Ruby JG, Stark A, Johnston WK, Kellis M, Bartel DP, Lai EC. 2007b. Evolution, biogenesis, expression, and target predictions of a sub-

- stantially expanded set of *Drosophila* microRNAs. *Genome Res* **17**: 1850–1864.
- Ryder E, Ashburner M, Bautista-Llacer R, Drummond J, Webster J, Johnson G, Morley T, Chan YS, Blows F, Coulson D, et al. 2007. The DrosDel deletion collection: A *Drosophila* genomewide chromosomal deficiency resource. *Genetics* **177**: 615–629.
- Saito K, Ishizuka A, Siomi H, Siomi MC. 2005. Processing of pre-microRNAs by the Dicer-1–Loquacious complex in *Drosophila* cells. *PLoS Biol* **3**: e235. doi: 10.1371/journal.pbio.0030235.
- Stark A, Brennecke J, Russell RB, Cohen SM. 2003. Identification of *Drosophila* microRNA targets. *PLoS Biol* **1**: e60. doi: 10.1371/journal.pbio.0000060.
- Stark KL, Xu B, Bagchi A, Lai WS, Liu H, Hsu R, Wan X, Pavlidis P, Mills AA, Karayiorgou M, et al. 2008. Altered brain microRNA biogenesis contributes to phenotypic deficits in a 22q11-deletion mouse model. *Nat Genet* **40**: 751–760.
- Stowers RS, Schwarz TL. 1999. A genetic method for generating *Drosophila* eyes composed exclusively of mitotic clones of a single genotype. *Genetics* **152**: 1631–1639.
- Tomari Y, Matranga C, Haley B, Martinez N, Zamore PD. 2004. A protein sensor for siRNA asymmetry. *Science* **306**: 1377–1380.
- Tomari Y, Du T, Zamore PD. 2007. Sorting of *Drosophila* small silencing RNAs. *Cell* **130**: 299–308.
- Triboulet R, Chang HM, Lapierre RJ, Gregory RI. 2009. Post-transcriptional control of DGCR8 expression by the Microprocessor. *RNA* **15**: 1005–1011.
- Vaucheret H, Vazquez F, Cr  t   P, Bartel DP. 2004. The action of ARGONAUTE1 in the miRNA pathway and its regulation by the miRNA pathway are crucial for plant development. *Genes Dev* **18**: 1187–1197.
- Vaucheret H, Mallory AC, Bartel DP. 2006. AGO1 homeostasis entails coexpression of *MIR168* and *AGO1* and preferential stabilization of miR168 by AGO1. *Mol Cell* **22**: 129–136.
- Williamson WR, Wang D, Haberman AS, Hiesinger PR. 2010. A dual function of V0-ATPase a1 provides an endolysosomal degradation mechanism in *Drosophila melanogaster* photoreceptors. *J Cell Biol* **189**: 885–899.
- Wu H, Xu H, Miraglia LJ, Crooke ST. 2000. Human RNase III is a 160-kDa protein involved in preribosomal RNA processing. *J Biol Chem* **275**: 36957–36965.
- Xie Z, Kasschau KD, Carrington JC. 2003. Negative feedback regulation of *Dicer-like1* in *Arabidopsis* by microRNA-guided mRNA degradation. *Curr Biol* **13**: 784–789.
- Yang JS, Lai EC. 2011. Alternative miRNA biogenesis pathways and the interpretation of core miRNA pathway mutants. *Mol Cell* **43**: 892–903.
- Yang JS, Maurin T, Robine N, Rasmussen KD, Jeffrey KL, Chandwani R, Papapetrou EP, Sadelain M, O’Carroll D, Lai EC. 2010. Conserved vertebrate *mir-451* provides a platform for Dicer-independent, Ago2-mediated microRNA biogenesis. *Proc Natl Acad Sci* **107**: 15163–15168.
- Yi R, Qin Y, Macara IG, Cullen BR. 2003. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes Dev* **17**: 3011–3016.
- Zdanowicz A, Thermann R, Kowalska J, Jemielity J, Duncan K, Preiss T, Darzynkiewicz E, Hentze MW. 2009. *Drosophila* miR2 primarily targets the m<sup>7</sup>GpppN cap structure for translational repression. *Mol Cell* **35**: 881–888.
- Zhou R, Hotta I, Denli AM, Hong P, Perrimon N, Hannon GJ. 2008. Comparative analysis of Argonaute-dependent small RNA pathways in *Drosophila*. *Mol Cell* **32**: 592–599.